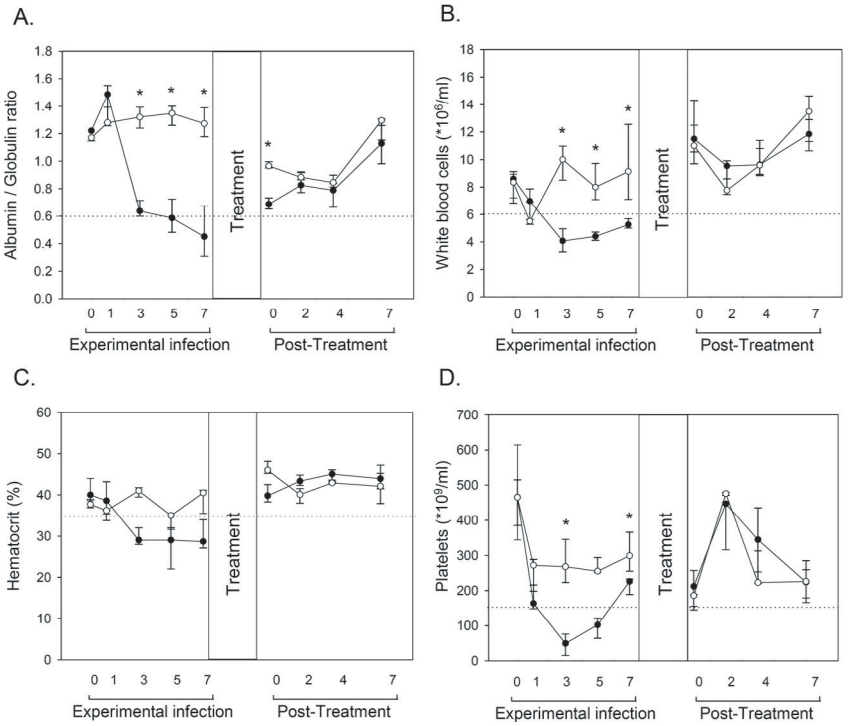
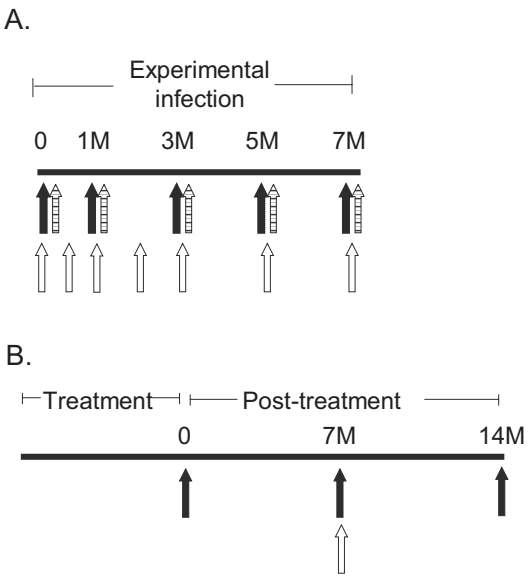
Online Materials



Supplementary Figure 1. Laboratory results during the experimental infection and following treatment. Experimentally infected dogs (closed circles) and uninfected controls (open circles) were evaluated for the albumin to globulin ratio (panel A), WBC count (panel B), hematocrit (panel C) and platelet count (panel D), during the experimental infection and following treatment. Values that are different from uninfected control dogs are indicated ($p < 0.05$ *). The dashed horizontal line indicates the lower normal value.

Supplementary Table I. Laboratory results of *L. infantum* naturally infected dogs.

	Albumin/Globulin ratio	Hematocrit (%)	WBC ($\times 10^3/\mu L$)	PLT ($\times 10^6/\mu L$)	Urea (mg/dL)	Creatinine (mg/dL)
Median (25th–75th percentile)	0.35 (0.23–0.41)	30 (29–40)	7.02 (6.4–7.7)	146 (105–219)	39.5 (26–50)	0.83 (0.72–1.2)
Normal values	> 0.6	35–55	6–17	150–700	11–70	0.36–1.36



Supplementary Figure 2. Sampling scheme during the experimental infection and following treatment. Spleen aspirates were obtained from experimentally-infected dogs (black arrows) and uninfected controls (striped arrows) at various time points during the experimental infection (panel A). Experimentally-infected dogs were further sampled at various time points after the treatment was stopped (panel B). Serum samples for anti-*Leishmania* total IgG and IgG subclass ELISA were obtained from the experimentally-infected dogs during the infection and after the treatment (clear arrows; panels A, B).

Supplementary Table II. Cytokine, transcription factor and chemokine primers sequences used for real-time qPCR.

Target	Forward primer (5' to 3')	Reverse primer (3' to 5')	GenBank reference sequence	$MgCl_2$ (mM)	Primers (nM)
IFN-γ	394F - AAGGAAGACATGCTTGGCAAG	487R- CCTGCAGATCGTTCACAGGAA	MN_001003174	3	50/50[a]
TNF-α	290F - AGCAAACCCCGAAGCTGAG	392R- CGGCACTATCAGCTGGTTGTC	S74068	3	100/100
IL-5	70F - TGCCTATGTTTCTGCCTTTGCT	180F - ATCAGGTTCCCATCGCCTATC	AF331919	3	50/50
IL-4	112F - CTCACCAGCACCTTTGTCCA	202R- AGTCGTTTCTCGCTGTGAGGA	MN_001003159	3.5	100/100
IL-10[b]	304F - CCACGACCCAGACATCAAGAA	406R - ACAGGGAAGAAATCGGTGACA	U33843	3.5	100/100
TGF-β	720F - CCACTGTTCCTGTGACAGCAA	823R- GTCGGTTCATGCCATGAATG	L34956	2.5	100/100
β−actin	577F - CTCACGGAGCGTGGCTACA	661R-GGCGACGTAGCAGAGCTTCTT	AF021873	3.5	100/100
T-bet	1465F-CCCCTTCGGTGTGGACTGAGA	1585R- GGAGGAGCTGTCGCCACTGG	AF241243	3.5	50/50
GATA-3	61F-CCCAAGAGCAGCTCGTTCA	166R- GCGTTGGAGTGGTCAGCAT	AF459800	3	100/100
IP-10	123F-TGAAATGATTCCTGCAAGTCCAT	193R- TCTCCCCACTCTTTTTCATTGTG	[c]	3	50/50
RANTES	113F-GCCTCTGCCTCCCCATATG	173R- GGCCGGAAATGTAGGCAAA	MN_1003010.1	3.5	50/50
MIP-1α	85F- ACCCCAATAGCCTGGTGCTT	143R- CGACTATGAACTTGCGTGGAATC	MN_1005251.1	3.5	50/50
MCP-1	69F- CCCAGCCAGATGCAATTATTTC	149R- TGGCCAGCCTCTGAATTGAG	MN_1003297.1	3.5	50/50

[a] Forward and reverse primer concentrations.
[b] Primer sequences were kindly provided by Dr Reinhard K. Straubinger, Institute of Immunology, University of Leipzig.
[c] Frangogiannis et al. [17].